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<p>(54) Title: PHARMACOLOGICAL AGENTS</p> <p>(57) Abstract</p> <p>Antagonists selective for the GluR5 receptor are useful for the treatment of pain. Also disclosed are novel decahydroisoquinoline derivatives which are selective GluR5 receptor antagonists.</p>			

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Pharmacological Agents

The present invention relates to a new class of glutamate receptor antagonists useful for the treatment of 5 pain.

L-Glutamate mediates excitatory neurotransmission in the mammalian central nervous system through its action at glutamate receptors. There are two broad classes of glutamate receptors, known as the ionotropic glutamate 10 receptor and the metabotropic glutamate receptor. Within the class of ionotropic glutamate receptor are three classes, known as the N-methyl-D-aspartate (NMDA), (R,S)-2-amino-3-(3-hydroxy-5-methyl-isoxazol-4-yl)propanoate (AMPA) and kainate (KA) receptors. Molecular biological studies 15 have established that AMPA receptors are composed of subunits (GluR1-4) that can assemble to form functional channels. Five kainate receptors, classified as either high affinity (KA1 and KA2) or low affinity (GluR5, GluR6 and GluR7) kainate receptors have been identified. (Bleakman et 20 al, *Molecular Pharmacology*, 1996, Vol. 49, No. 4, pgs. 581-585).

European Patent Application Publication No. 590789A1 and United States Patent No. 5,446,051 disclose that certain decahydroisoquinoline derivatives are AMPA receptor 25 antagonists, and as such are useful in the treatment of many different conditions, including pain. There is no disclosure of any compound actually being tested for use in the treatment of pain.

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Surprisingly, it has now been found that a compound within the scope of European Patent Application Publication No. 590789A1, namely (3S,4aR,6S,8aR)-6-[(1(2)H-tetrazol-5-yl)methoxymethyl]decahydroisoquinoline-3-carboxylic acid, is 5 a selective GluR5 antagonist and is effective in animal models of pain. It is therefore believed that a new pharmacological class of agents, represented by (3S,4aR,- 6S,8aR)-6-[(1(2)H-tetrazol-5-yl)methoxymethyl]decahydro-isoquinoline-3-carboxylic acid, has been found for the 10 treatment of pain.

According to one aspect, therefore, the present invention provides a method for the treatment of pain, which comprises administering to a mammal in need of treatment an effective amount of a selective GluR5 receptor antagonist.

15 According to another aspect, the present invention provides the use of a selective GluR5 receptor antagonist for the manufacture of a medicament for the treatment of pain.

GluR5 receptor antagonists may be identified by 20 radiolabelled ligand binding studies at the cloned and expressed human GluR5 receptor (Korczak et al., 1994, Recept. Channels 3; 41-49), by whole cell voltage clamp electro-physiological recordings of functional activity at the human GluR5 receptor (Korczak et al., 1994, Recept. 15 Channels 3; 41-49) and by whole cell voltage clamp electrophysiological recordings of currents in acutely isolated rat dorsal root ganglion neurons (Bleakman et al., 1996, Mol. Pharmacol. 49; 581-585).

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The selectivity of compounds acting at GluR5 receptors may be determined by measurement of activity at other AMPA and kainate receptors including receptor-ligand binding studies and whole-cell voltage clamp electro-physiological 5 recordings of functional activity at human GluR1, GluR2, GluR3 and GluR4 receptors (Fletcher et al., 1995, *Recept. Channels* 3; 21-31), receptor-ligand binding studies and whole-cell voltage clamp electrophysiological recordings of functional activity at human GluR6 receptors (Hoo et al., 10 *Recept. Channels* 2; 327-338) and whole-cell voltage clamp electrophysiological recordings of functional activity at AMPA receptors in acutely isolated cerebellar Purkinje neurons (Bleakman et al., 1996, *Mol. Pharmacol.* 49; 581-585) and other tissues expressing AMPA receptors (Fletcher and 15 Lodge, 1996, *Pharmacol. Ther.* 70; 65-89).

Preferably, the selective GluR5 receptor antagonist has a binding affinity of at least 10 fold greater for GluR5 than that for other glutamate receptors, more preferably at least 100 fold greater.

20 The selective GluR5 antagonist for use according to the invention may be a single compound or combination of compounds capable of functioning as an antagonist that is selective for the GluR5 receptor over other ionotropic glutamate receptors. For example, it may be a combination 25 of a compound capable of functioning as an antagonist at the GluR5 receptor and one or more other glutamate receptors in combination with one or more compounds capable of blocking its actions at the one or more other ionotropic glutamate

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receptors. Preferably, the selective GluR5 antagonist is a single compound.

The following compounds have been found to be selective GluR5 receptor antagonists and are therefore preferred for 5 use according to the invention: 3SR,4aRS,6SR,8aRS-6-(((1*H*- tetrazole-5-yl)methyl)oxy)methyl)-1,2,3,4,4a,5,6,7,8,8a- decahydroisoquinoline-3-carboxylic acid, 3S,4aR,6S,8aR-6- (((1*H*-tetrazole-5-yl)methyl)oxy)methyl)-1,2,3,4,4a,5,6,- 7,8,8a-decahydroisoquinoline-3-carboxylic acid,
10 3SR,4aRS,6SR,8aRS-6-((4-carboxy)phenyl)methyl)-1,2,3,4,- 4a,5,6,7,8,8a-decahydroisoquinoline-3-carboxylic acid and 3S,4aR,6S,8aR-6-((4-carboxy)phenyl)methyl)-1,2,3,4,4a,- 5,6,7,8,8a-decahydroisoquinoline-3-carboxylic acid.

The results of evaluating the activity of the 15 aforementioned decahydroisoquinoline derivatives at the GluR5 receptor and other ionotropic glutamate receptors in the tests described above are given in Tables 1 and 2 below.

TABLE 1

20 Selectivity Profile for Compounds
of the Invention in Binding Studies

Cell lines (HEK293 cells) stably transfected with human GluR receptors were employed. Displacement of ³H AMPA by increasing concentrations of test compound was used on GluR1- 25 4-expressing cells and ³H kainate (KA) on GluR5 and expressing cells. Estimated activity (Ki) in nM was as follows.

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Test Compound	GluR1	GluR2	GluR3	GluR4	GluR5	GluR6
A*	150686± 49789 (3)	35337± 6163 (3)	47793± 8770 (3)	31606± 5914 (3)	3061± 1038 (3)	6±5% disp1 @ 100µM (3)
B*	--	--	--	--	5823 (2)	--
C*	21% at 1mM	55% at 1mM	--	23% at 1mM	6810	--

A - 3SR, 4aRS, 6SR, 8aRS-6-[(1(2)H-tetrazol-5-yl)methoxy-methyl]decahydroisoquinoline-3-carboxylic acid
(average of 3 results)

5

B - Compound of Example 1 (average of 2 results)

C - 3SR, 4aRS, 6SR, 8aRS-6-(((4-carboxy)phenyl)methyl)-1,2,3-
4,4a,5,6,7,8,8a-decahydroisoquinoline-3-carboxylic acid
(one result only)

D - Compound of Example 2
15 -- Not tested

TABLE 2
Selectivity Profile for Compounds
of the Invention in Electrophysiological Studies

Functional studies were carried out on HEK293 cells
20 stably transfected with human GluR receptors and on acutely isolated dorsal root ganglion neurons (DRG) using patch-clamp technology (Bleakman *et al.*, 1996, *Mol. Pharmacol.*, 49, 581-585). IC50 values (µM) for test Compound A were estimated for GluR1-4 vs 100 µM AMPA and GluR5 and GluR6 vs 25 100 µM KA with the following results:

Test Compound	GluR1	GluR2	GluR3	GluR4	GluR5	GluR6	DRG*
A	>100	>100	>100	>100	3.9±0.5	>100	0.60±0.06
D	--	--	--	>100	--	>100	0.98±0.07

*Based on % inhibition of 30 µM kainate induced current.

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The forms of pain which may be treated according to the invention include severe, chronic, intractable and neuropathic pain.

The compounds 3S,4aR,6S,8aR-6-(((1H-Tetrazole-5-yl)methyl)oxy)methyl)1,2,3,4,4a,5,6,7,8,8a-decahydro-isoquinoline-3-carboxylic acid; and 3S,4aR,6S,8aR-6-(((4-carboxy)phenyl)methyl)1,2,3,4,4a,5,6,7,8,8a-decahydro-isoquinoline-3-carboxylic acid and their pharmaceutically acceptable salts are believed to be novel and are provided as a further aspect of the invention. The invention also provides a pharmaceutical composition comprising one of these compounds and a pharmaceutically acceptable diluent or carrier. They may be prepared, and formulated into pharmaceutical compositions, by the general methods described in European Patent Application No. 590789A1 and United States Patent No. 5,446,051.

The ability of selective GluR5 receptor antagonists to treat pain in mammals may be demonstrated using the well known formalin, tail flick and acetic acid-induced writhing tests.

1) formalin test

Male Sprague-Dawley rats (200-250g; Charles River, Portage, MI) were housed in group cages and maintained in a constant temperature and a 12h light/12h dark cycle 4-7 days before the studies were performed. Animals had free access to food and water at all times prior to the day of the experiment.

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Drugs or vehicles were administered intraperitoneally (i.p.) or orally (p.o.) by gavage in a volume of 1 ml/kg.

The test was performed in custom made Plexiglas® boxes 25x25x20x cm in size (according to Shibata et al., Pain 38; 5 347-352, 1989, Wheeler-Aceto et al., Pain, 40; 229-238, 1990). A mirror placed at the back of the cage allowed the unhindered observation of the formalin injected paw. Rats were acclimated individually in the cubicles at least 1 hour prior to the experiment. All testing was conducted between 10 08:00 and 14:00 h and the testing room temperature was maintained at 21-23°C. Test compounds were administered 30 minutes prior to the formalin injection. Formalin (50 µl of a 5% solution in saline) was injected subcutaneously into the dorsal lateral surface of the right hind paw with a 27 15 gauge needle. Observation started immediately after the formalin injection. Formalin-induced pain was quantified by recording in 5 minute intervals the number of formalin injected paw licking events and the number of seconds each licking event lasted. These recordings were made for 50 20 minutes after the formalin injection.

Different scoring parameters have been reported for the formalin test. The total time spent licking and biting the injected paw was demonstrated to be most relevant (Coderre et al., *Eur. J. Neurosci.* 6; 1328-1334, 1993; Abbott et al., 25 Pain, 60; 91-102, 1995) and was chosen for the testing score. The early phase score is the sum of time spent licking in seconds from time 0 to 5 minutes. The late phase was scored in 5 minute blocks from 15 minutes to 40 minutes

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and is expressed accordingly or also by adding the total number of seconds spent licking from minute 15 to minute 40 of the observation period. Data are presented as means with standard errors of means (\pm SEM). Data were evaluated by 5 one-way analysis of variance (ANOVA) and the appropriate contrasts analyzed by Dunnett "t" test for two sided comparisons. Differences were considered to be significant if the P-value was less than 0.05 and indicated by asterisk. Statistics were determined at the 5 minute time point and at 10 5 minute intervals between 15 and 40 minutes. Where data are expressed as total amount of time spent licking in the late phase, statistics were performed on the total time spent licking as well and are indicated accordingly.

In this test, the compound of Example 1 was found to be 15 active at doses in the range 10 to 100 mg/kg in reducing second phase paw licking time.

2) Tail flick test

This well known test measures the effect of a test compound on the time taken for an animal to flick its tail 20 out of a focused beam of light. In the test, a beam of light from a lamp is focused on a surface, and the lamp is then switched off. Treated and untreated (control) animals are restrained, with their tails positioned at the focal point of the beam of light from the lamp. The lamp is then 25 switched on, and the time taken for the animal's tail to respond by moving is recorded.

Subcutaneous administration of the compound of Example 1 in mice at doses of 3, 10, and 30 mg/kg produced a dose

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dependent increase in response time. Oral administration of the same compound at doses of 0.03, 0.1, 0.3 and 3 mg/kg to cynomolgous primates also produced a dose dependent increase in response time. These data show that the compound of 5 Example 1, which is a selective GluR5 antagonist, is effective at treating pain, and has unexpected oral activity in cynomolgous primates.

3) Acetic acid induced mouse writhing test.

This test measures the ability of a test compound to 10 reduce the amount of writhing induced by intraperitoneal injection of acetic acid in mice.

Doses of test compound or control are administered to male CD-1 mice. Each animal is then administered 0.5% acetic acid in a volume of 0.01 mg/g intraperitoneally. The 15 animals are then placed in individual plexiglas observation chambers and the total number of writhes (flattening of the abdominal wall and asymmetric stretching and extending of the body and hindlimbs) recorded between 5 and 10 minutes after acetic acid administration.

20 The compound of Example 1 was administered at a dose of 1, 3, 10 and 30 mg/kg, and produced a dose dependent reduction in writhing.

The particular dose of antagonist administered according to this invention will of course be determined by 25 the particular circumstances surrounding the case, including the activity of the particular antagonist administered, the route of administration, the particular condition being treated, and similar considerations. The antagonist can be

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administered by a variety of routes including oral, rectal, transdermal, subcutaneous, intravenous, intramuscular, or intranasal routes. Alternatively, the antagonist may be administered by continuous infusion. A typical daily dose 5 will contain from about 0.001 mg/kg to about 100 mg/kg of the antagonist. Preferably, daily doses will be about 0.05 mg/kg to about 50 mg/kg, more preferably from about 0.1 mg/kg to about 20 mg/kg.

The following examples illustrate the preparation of 10 novel compounds that are selective GluR5 antagonists.

Tetrahydrofuran was dried by distillation from sodium. All other solvents and reagents were used as obtained. The reactions were generally monitored for completion using thin layer chromatography (TLC). Thin layer chromatography was 15 performed using E. Merck Kieselgel 60 F₂₅₄ plates, 5 cm X 10 cm, 0.25 mm thickness. Spots were detected using a combination of UV and chemical detection (plates dipped in a ceric ammonium molybdate solution [75 g of ammonium molybdate and 4 g of cerium (IV) sulfate in 500 mL of 10% 20 aqueous sulfuric acid] and then heated on a hot plate).

Elemental analyses for carbon, hydrogen and nitrogen were determined on a Control Equipment Corporation 440 Elemental Analyzer. "Chromatography" refers to flash chromatography (Still, WC; Kahn, M; Mitra, A. *J. Org. Chem.* 1978, 43, 25 2923) on 230-400 mesh Silica Gel 60, using the amount of silica gel and solvent of elution referred to parenthetically in the text. "Cation exchange chromatography" refers to ion exchange with Dowex 50X-8

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(100-200) resin (H⁺ form). The resin was prepared by washing (in a coarse porosity sintered glass funnel) with water, then methanol, then water, then 3N ammonium hydroxide (pH \geq 12), then water, then 1N HCl (pH \leq 1), then water 5 until the pH is neutral. The resin was packed into a glass column in water, and the compound (which is dissolved in water at a pH between 2 and 7) was slowly eluted onto the resin, then the column washed with water until the pH is neutral, then 50% aqueous THF, then water. The compound is 10 eluted off of the resin with 10% aqueous pyridine, and product containing fractions (which are detected with ninhydrin stain on a TLC plate) are combined and concentrated in vacuo. Water is added and the mixture concentrated in vacuo. This procedure is repeated two more 15 times, and ensures complete removal of pyridine.

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EXAMPLE 1

*3S,4aR,6S,8aR-6-(((1*H*-Tetrazole-5-yl)methyl)oxy)methyl)-1,2,3,4,4a,5,6,7,8,8a-decahydroisoquinoline-3-carboxylic Acid*

5

A. Ethyl *3S,4aR,6S,8aR* 6-(((Cyanomethyl)oxy)methyl)-2-methoxycarbonyl-1,2,3,4,4a,5,6,7,8,8a-decahydroisoquinoline-3-carboxylate: A solution of 12.8 g (42.6 mmol) of ethyl *3S,4aR,6S,8aR*-6-hydroxymethyl-2-methoxycarbonyl-1,2,3,4,-10 *4a,5,6,7,8,8a*-decahydroisoquinoline-3-carboxylate (Ornstein, et al., *Journal of Organic Chemistry*, 1994, 59, 7862-7869), 10.6 g (85.3 mmol) of ((methoxy)ethoxy)methyl chloride 16.5 g (127.9 mmol) of *N,N*-diisopropylethylamine and 20 mg of 4-*N,N*-dimethylaminopyridine in 70 mL of methylene chloride was 15 heated to reflux for four hours, then cooled, diluted with 150 mL of ether and washed twice with 100 mL each of 10% aqueous sodium bisulfate. The organic phase was dried (magnesium sulfate), filtered, and concentrated *in vacuo*. The residual oil was dissolved in 70 mL of methylene and 20 treated with 17.0 mL (12.7 g, 127.9 mmol) of trimethyl-silyl cyanide, then the solution was cooled to 0°C and treated with 1.3 mL (1.5 g, 10.7 mmol) of boron trifluoride etherate. The resulting solution was allowed to warm to room temperature. After three hours at room temperature, 25 the reaction mixture was treated with 100 mL of 10% aqueous potassium carbonate, then 150 mL of ether. The phases were separated and the organic phase washed one more time with 100 mL of 10% aqueous potassium carbonate. The organic

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phase was dried (magnesium sulfate), filtered, and concentrated in vacuo. Chromatography of the residue (350 g of silica gel, 30% ethyl acetate/hexane) afforded 11.9 g (83%) of the title compound.

5 Analysis calculated for $C_{17}H_{26}N_2O_5$: C, 60.34; H, 7.74; N, 8.28. Found: C, 60.06; H, 7.69; N, 8.31.
[α]_D = -33.5° (c = 1, CH_2Cl_2).

B. A mixture of 11.7 g (34.6 mmol) of the compound from
10 Example 1A and 23.0 g (69.2 mmol) of tributyltin azide was heated to 100°C for five days. The mixture was treated with 100 mL of 6 N hydrochloric acid, and heated continued at 100°C. After about 18 hours, the reaction mixture was allowed to cool to room temperature, then extracted with 50 mL of ether, 50 mL of dichloromethane and 50 mL of ether, then the aqueous phase was concentrated in vacuo. Cation exchange chromatography of the residue afforded a solid that was suspended in acetone, refluxed for one hour, then filtered and washed with acetone and ether, and dried in
15 vacuo at 60°C to afford 8.5 g (83%) of the title compound.
Analysis calculated for $C_{13}H_{21}N_5O_3 \cdot 0.33 H_2O \cdot 0.33 C_3H_6O$: C, 52.43; H, 7.44; N, 21.84. Found: C, 52.74; H, 7.22; N, 21.50.
[α]_D = -21.6° (c = 1, 1N HCl).

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EXAMPLE 2

Preparation of 3*S*,4*aR*,6*S*,8*aR*-6-(((4-carboxy)phenyl)methyl)-1,2,3,4,4*a*,5,6,7,8,8*a*-decahydroisoquinoline-3-carboxylic acid

5

A. Methyl 4-(diethylphosphonomethyl)benzoate: A solution of 25.0 g (110 mmol) of methyl 4-bromomethylbenzoate and 37 mL (36.3 g, 220 mmol) of triethyl phosphite in 150 mL of toluene was heated for 18 hours at reflux, then cooled and concentrated *in vacuo*. Chromatography (400 g of silica gel, ethyl acetate) of the residue afforded 30.6 g (98%) of the title compound.

B. Ethyl 3*S*,4*aR*,6*S*,8*aR*-6-(((4-methoxycarbonyl)phenyl)-methyl)-1,2,3,4,4*a*,5,6,7,8,8*a*-decahydroisoquinoline-3-carboxylate: 14.1 g (49.4 mmol) of the compound from Example 2A and 48 mL (of a 1.0 M solution) of sodium bis(trimethyl-silyl)amide in 100 mL of THF was stirred 45 min at 0°C, then treated with 10.0 g of ethyl 3*S*,4*aR*,8*aR*-6-oxo-2-methoxy-carbonyl-1,2,3,4,4*a*,5,6,7,8,8*a*-decahydroisoquinoline-3-carboxylate in 40 mL of THF. After 15 minutes at 0°C, the reaction was quenched with 100 mL of water and extracted three times with 150 mL each of ether. The combined organics were dried (magnesium sulfate), filtered and concentrated *in vacuo*. The residue was dissolved in 500 mL of ether, treated with 3.0 g of 5% palladium on carbon, and hydrogenated at room temperature and one atmosphere of hydrogen for 24 hours. The mixture

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was diluted with 500 mL of ether, filtered through a pad of diatomaceous earth, and the filtrate concentrated in vacuo. Chromatography (400 g of silica gel, 25% ethyl acetate/hexane) of the residue afforded 12.0 g (81%) of the 5 title compound, as a mixture of methyl and ethyl esters from transesterification of the methyl ester during hydrogenation.

C: 12.0 g of the compound from Example 2B was heated to 10 reflux for 18 hours with 100 mL of 6N hydrochloric acid, then cooled to room temperature. The resulting solid was filtered, washing with water, acetone and ether, and dried in vacuo at 60°C to afford 6.2 g (57%) of the title compound.

15 Analysis calculated for $C_{18}H_{23}NO_4 \cdot HCl \cdot 1.25 H_2O$: C, 57.44; H, 7.10; N, 3.72. Found: C; 57.44; H, 6.69; N, 3.76. $[\alpha]_D = -4.8^\circ$ (c = 1, 1N HCl).

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We claim:

1. A method for the treatment of pain, which comprises
5 administering to a mammal in need of treatment an
effective amount of a selective GluR5 receptor
antagonist.
2. A method as claimed in Claim 1, in which the selective
10 GluR5 receptor antagonist is selected from 3SR,4aRS,-
6SR,8aRS-6-(((1*H*-tetrazole-5-yl)methyl)oxy)methyl)-
1,2,3,4,4a,5,6,7,8,8a-decahydroisoquinoline-3-
carboxylic acid, 3*S*,4a*R*,6*S*,8a*R*-6-(((1*H*-tetrazole-5-
15 yl)methyl)oxy)methyl)1,2,3,4,4a,5,6,7,8,8a-
decahydroisoquinoline-3-carboxylic acid,
3*S**R*,4*a**R**S*,6*S**R*,8*a**R**S*-6-((4-carboxy)phenyl)methyl)-
1,2,3,4,4a,5,6,7,8,8a-decahydroisoquinoline-3-
20 carboxylic acid and 3*S*,4*a**R*,6*S*,8*a**R*-6-((4-carboxy)-
phenyl)methyl)1,2,3,4,4a,5,6,7,8,8a-decahydro-
isoquinoline-3-carboxylic acid.
3. The use of a selective GluR5 receptor antagonist for
the manufacture of a medicament for the treatment of
25 pain.
4. Use as claimed in Claim 3, in which the selective GluR5
receptor antagonist is selected from 3*S**R*,4*a**R**S*,
6*S**R*,8*a**R**S*,-6-(((1*H*-tetrazole-5-yl)methyl)oxy)methyl)-

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1,2,3,4,4a,5,6,7,8,8a-decahydroisoquinoline-3-carboxylic acid, 3S,4aR,6S,8aR-6-(((1H-tetrazole-5-yl)methyl)oxy)methyl)1,2,3,4,4a,5,6,7,8,8a-decahydroisoquinoline-3-carboxylic acid, 3SR,4aRS,6SR,8aRS-6-((4-carboxy)phenyl)methyl)-1,2,3,4,4a,5,6,7,8,8a-decahydroisoquinoline-3-carboxylic acid and 3S,4aR,6S,8aR-6-((4-carboxy)phenyl)methyl)-1,2,3,4,4a,5,6,7,8,8a-decahydroisoquinoline-3-carboxylic acid.

10

5. A compound which is selected from 3S,4aR,6S,8aR-6-(((1H-tetrazole-5-yl)methyl)oxy)methyl)-1,2,3,4,4a,5,6,7,8,8a-decahydroisoquinoline-3-carboxylic acid and 3S,4aR,6S,8aR-6-((4-carboxy)phenyl)methyl)1,2,3,4,4a,5,6,7,8,8a-decahydroisoquinoline-3-carboxylic acid.

15

6. A pharmaceutical composition, which comprises a compound as claimed in Claim 5 and a pharmaceutically acceptable diluent or carrier.

20

7. A pharmaceutical composition for the treatment of pain, which comprises a selective GluR5 receptor antagonist and a pharmaceutically acceptable diluent or carrier.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US98/06905

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) :C07D 217/06; A61K 31/47
US CL :514/307; 546/22, 147

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 514/307; 546/22, 147

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
noneElectronic data base consulted during the international search (name of data base and, where practicable, search terms used)
APS AND CAS ONLINE: pain? with glur5, ?decahydroisoquinolin?

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 5,446,051 A (ORNSTEIN) 29 August 1995, col. 1, line 66 - col. 3, line 11.	1-7

 Further documents are listed in the continuation of Box C. See patent family annex.

* Special categories of cited documents.	*T*	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance		
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O document referring to an oral disclosure, use, exhibition or other means	*&*	document member of the same patent family
P document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search 04 JUNE 1998	Date of mailing of the international search report 17 JUL 1998
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230	Authorized officer KIMBERLY JORDAN Telephone No. (703) 308-1235

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